

Thaw and Culture Details

Cell Line Name	PENN020i-588-6						
WiCell Lot Number	DB36448						
Provider	University of Pennsylvania – Dr. Daniel Rader						
Banked By	Penn Institute for Regenerative Medicine iPS Core Facility						
Thaw and Culture Recommendations	WiCell recommends thawing 1 vial into 1 well of a 6 well plate. WiCell recommends thawing using ROCK Inhibitor for best results.						
Culture Platform	Feeder Dependent						
	Medium: hESC Medium (KOSR)						
	Matrix: MEF						
Protocol	WiCell Feeder Dependent Protocol						
Passage Number	p13 These cells were cultured for 13 passages prior to freeze and post colony picking. Therefore, plated cells at thaw should be labeled passage 14.						
Date Vialed	16-March-2015						
Vial Label	iPS-588 SeV6 p13 03/16/15 KS						
Biosafety and Use Information	Appropriate biosafety precautions should be followed when working with these cells. The end user is responsible for ensuring that the cells are handled and stored in an appropriate manner. WiCell is not responsible for damages or injuries that may result from the use of these cells. Cells distributed by WiCell are intended for research purposes only and are not intended for use in humans.						

Testing Performed by WiCell

Test Description	Test Provider	Test Method	Test Specification	Result			
Karyotype by G-banding	WiCell	SOP-CH-003	Expected karyotype	See Report			
Post-Thaw Viable Cell Recovery	WiCell	SOP-CH-305	Recoverable attachment after passage	Pass			
Identity by STR	UW Translational Research Initiatives in Pathology Laboratory	PowerPlex 16 HS System by Defines profile		Pass			
Sterility	Steris	ST/07	Negative	Pass			
Mycoplasma	WiCell	SOP-QU-004	Negative	Pass			

Testing Reported by Provider

The Provider stated that some or all of the additional analyses listed below may have been performed for this cell line. For more information, publication and dbGaP links, where available, are provided on the cell line specific web page on the WiCell website.

- SNP microarray
- Flow Cytometry (Tra1-60 and SSEA-4)
- Differentiation into hepatocytes
- Infinium® Expanded Multi-Ethnic Genotyping Array (MEGAEX)



Approval Date	Quality Assurance Approval			
23-June-2016	5,19,2018 X JKG IKC Quality Assurance Signed by Gay, Jenna			



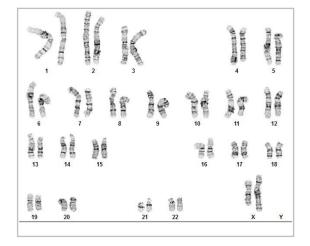
Chromosome Analysis Report: 071653

Date Reported: Thursday, May 10, 2018
Cell Line: PENN020i-588-6-DB36448 13639

Passage#: 16

Date of Sample: 5/7/2018 Specimen: Human IPS

Results: 46,XX



Cell Line Gender: Female

Reason for Testing: lot release testing

Investigator: WiCell

Cell: 34 Slide: G01

Slide Type: Karyotype

Total Counted: 20
Total Analyzed: 8

Total Karyogrammed: 4
Band Resolution: 500 - 525

Interpretation:

This is a normal karyotype; no clonal abnormalities were detected at the stated band level of resolution.

Completed by: Reviewed and Interpreted by:

CG(ASCP)
PhD, FACMG

A signed copy of this report is available upon request.

Date:	Sent By:	Sent To:	QC Review By:

Limitations: This assay allows for microscopic visualization of numerical and structural chromosome abnormalities. The size of structural abnormality that can be detected is >3-10Mb, dependent upon the G-band resolution obtained from this specimen. For the purposes of this report, band level is defined as the number of G-bands per haploid genome. It is documented here as "band level", i.e., the range of bands determined from the four karyograms in this assay. Detection of heterogeneity of clonal cell populations in this specimen (i.e.,mosaicism) is limited by the number of metaphase cells examined, documented here as "# of cells counted".

This assay was conducted solely for listed investigator/institution. The results may not be relied upon by any other party without the prior written consent of the Director of the WiCell Cytogenetics Laboratory. The results of this assay are for research use only. If the results of this assay are to be used for any other purpose, contact the Director of the WiCell Cytogenetics Laboratory.

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Short Tandem Repeat Analysis

HISTOLOGY - IHC - MOLECULAR - IMAGING

Department of Pathology and Laboratory Medicine TRIP Laboratory (Molecular)

http://www.pathology.wisc.edu/research/trip

info@wicell.org (888) 204-1782

Sample Report:

13639-STR

Sample Name on Tube: 13639-STR

 $37.0 \text{ ng/}\mu\text{L}$, (A260/280=1.86)

Sample Type: Cells

Cell Count: ~2 million cells

Requestor:

WiCell Research Institute

Quality Department

Sample Date: N/A **Receive Date:** 05/07/18 **Assay Date:** 05/09/18

File Name: STR 180510 wmr

Report Date: 05/15/18

STR Locus	STR Genotype Repeat #	STR Genotype						
FGA	16–18,18.2,19,19.2,20,20.2,21,21.2,22, 22.2, 23, 23.2, 24, 24.2, 25, 25.2, 26–30, 31.2, 43.2, 44.2,45.2, 46.2	Identifying						
TPOX	6-13	information has						
D8S1179	7-18	been redacted to						
vWA	10-22	protect donor						
Amelogenin	X,Y	confidentiality. If more information						
Penta_D	2.2, 3.2, 5, 7-17							
CSF1PO	F1PO 6-15 6S539 5, 8-15							
D16S539								
D7S820	6-14	Support.						
D13S317	7-15							
D5S818	7-16							
Penta_E	5-24							
D18S51	18S51 8-10, 10.2, 11-13, 13.2, 14-27							
D21S11	24,24.2,25,25.2,26-28,28.2,29,29.2, 30, 30.2,31, 31.2,32,32.2,33,33.2, 34,34.2,35,35.2,36-38							
TH01	4-9,9.3,10-11,13.3							
D3S1358	12-20							

Results: Based on the 13639-STR cells submitted by WiCell QA dated and received on 05/07/18, this sample (Label on Tube: 13639-STR) defines the STR profile of the human stem cell line PENN020i-588-6 comprising 29 allelic polymorphisms across the 15 STR loci analyzed.

Interpretation: No STR polymorphisms other than those corresponding to the human PENN020i-588-6 stem cell line were detected and the concentration of DNA required to achieve an acceptable STR genotype (signal/ noise) was equivalent to that required for the standard procedure (~1 ng/amplification reaction) from human genomic DNA. This result suggests that the 13639-STR sample submitted corresponds to the PENN020i-588-6 stem cell line and was not contaminated with any other human stem cells or a significant amount of mouse feeder layer cells.

Sensitivity: Sensitivity limits for detection of STR polymorphisms unique to either this or other human stem cell lines is $\sim 2-5\%$.

X WMR \mathbf{X} RMB **Digitally Signed on** 05/15/18 **Digitally Signed on** 05/15/18 PhD, Director / Co-Director TRIP Laboratory, Molecular UWHC Molecular Diagnostics Laboratory / UWSMPH TRIP Laboratory

Native Product Sterility Report



SAMPLE #:

18041158

DATE RECEIVED:

18-Apr-18

TEST INITIATED:

20-Apr-18

TEST COMPLETED:

04-May-18

SAMPLE NAME / DESCRIPTION:

JFRBi1 WB66746 13644

JFNY1 WB66747 13645

UCSD216i-114-1 WB66784 13646 PENN020i-588-6 DB36448 13647 PENN070i-408-1 DB35065 13648 PENN068i-697-3 DB36611 13649 PENN091i-588-2 DB35156 13650 PENN135i-30-8 DB34725 13651 STAN004i-147-1 WB66789 13652 STAN057i-162-1 WB66790 13653

UNIQUE IDENTIFIER:

NA

PRODUCT REGISTRATION:

Other: Human iPS cells

TEST RESULTS:

WiCell

504 S Rosa Rd, Rm 101

Madison, WI 53719

# Tested	# Positives (Growth)	- Control
10	0	2 Negatives

TEST SUMMARY:

# Samples	Media Type	Volume (mL)	Incubation Temperature (° C)	Incubation Duration (Days)
10	TSB	40	20 - 25	14
10	FTG	40	30 - 35	14

REFERENCE:

Processed according to LAB-003: Sterility Test Procedure

METHOD VALIDATION / PD #:

000053

TEST METHODOLOGY:

USP - Direct Transfer

COMMENTS:

"Reported as" per packing slip

REVIEWED BY

DATE D7MAY18

Specific test results may not be indicative of the characteristics of any other samples from the same lot or similar lots. This test report shall not be reproduced, except in full, without prior written approval. Liability is limited to the costs of the tests.



Mycoplasma Detection Assay Report Testing Performed by WiCell

Testing Performed by WiCell Lot Release Testing April 16, 2018

FORM SOP-QU-004.01 Version G Edition 02 Reported by: AP Reviewed by: DF BD Monolight 180

		Read	ing A	A Reading B		ling B	В	Ratio		
#	Sample Name	RLU1	RLU2	Ave	RLU1	RLU2	Ave	B/A	Result	Comments/Suggestions
1	PENN020i-588-6-DB36448 13639	384	398	391	180	176	178	0.46	Negative	
2	Positive (+) Control	427	435	431	23689	23939	23814	55.25	Positive	
3	Negative (-) Control	669	659	664	80	83	81.5	0.12	Negative	

